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²⁷Al, ¹⁵N, ¹³C and ¹H NMR spectra and negative-ion electrospray mass spectra of the 2:1 aluminium(III) complexes of azo dyes derived from anthranilic acid

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Abstract

The ²⁷Al, ¹⁵N, ¹³C and ¹H NMR spectra in DMSO and mass spectra of 2:1 aluminium(III) complexes (1b–4b) derived from anthranilic acid azo coupling products with 4-tert.butylphenol (1a), 2-naphthol (2a) acetoacetanilide (3a) 3-methyl-1-phenylpyrazol-5-one (4a) were measured and analysed. It was found that the aluminium atom was six-coordinated, being bound to two oxygens and the nitrogen originating from anthranilic acid. \bigcirc 2001 Published by Elsevier Science Ltd.

Keywords: Aluminium(III)-azo dye complexes, ²⁷Al NMR; ¹⁵N NMR; ¹³C NMR; ¹H NMR; Negative-ion electrospray mass spectra

1. Introduction

Metal-dye complexes play a very important role in dyestuff technology [1-5]; chromium, cobalt, copper, nickel and aluminium derivatives are used most frequently for these purposes [1-6].

We have shown in previous papers [7–10] that NMR spectroscopy is a very sensitive probe to study such compounds. The aim of this paper was to characterise 2:1 aluminium(III) complexes of azo dyes **1b–4b** (Scheme 1) derived from anthranilic acid azo coupling products with 4-*tert*. butylphenol (**1a**), 2-naphthol (**2a**), acetoacetanilide (3a) and 3-methyl-1-phenylpyrazol-5-one (4a) (Scheme 2) by means of ²⁷Al, ¹⁵N, ¹³C and ¹H NMR and to study the structure from the view-point of coordination at the aluminium atom and to compare the data with those published for analogous complexes in which substituted 2-hydroxy-anilines were used in preparation of starting azo dyes [9] (i.e. with compounds in which –COO group was substituted by –O group).

2. Experimental

2.1. Synthesis

Ammonium salts of compounds **1b–3b** (Scheme 1) were prepared from appropriate precursors **1a–3a**

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1b

2b





(Scheme 2) and $Al_2(SO_4)_3 * 18H_2O$ and ammonium analogously to cobalt derivatives in formamide solution [5]. Compound **4b** was prepared in the following way: 1.28 g of **4a** was dissolved in 20 ml of formamide and barium hydroxide dissolved in 10 ml of formamide was added. The mixture was heated 20 min at 70 °C and solid barium salt of **4a** was formed. Then 1.59 g of $Al_2(SO_4)_3 * 18H_2O$ in 20 ml of formamide was added and the mixture was stirred overnight at 90 °C. The reaction mixture was poured into 30 ml of water and the raw product was dried over P_2O_5 .

All aluminium complexes were purified using column chromatography (Al₂O₃, methanol). Methanol was evaporated in vacuo at 25 °C.

At N_b ^{15}N labelled compounds were prepared analogously in formamide solution using Na

 $^{15}NO_2$ (95% ^{15}N) in preparation of compounds **1a–4a**.

2.2. NMR spectroscopy

The ²⁷Al (93.84 MHz), ¹⁵N (36.50 MHz), ¹³C (90.56 MHz) and ¹H (360.13 MHz) NMR spectra of compounds **1b–4b** were measured on a Bruker AMX 360 spectrometer equipped with 5 mm broadband probe and 5 mm broadband inverse probe and a Silicon Graphic Indy computer in hexadeuteriodimethyl sulfoxide at ambient temperature. The ²⁷Al chemical shifts are referred to external Al(OH)₆³⁺ in water [δ (²⁷Al)=0.0], ¹⁵N chemical shifts to external neat nitromethane [δ (¹⁵N)=0.0] and ¹³C and ¹H chemical shifts were referred to the central peak of DMSO-*d*₆



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 $[\delta(^{13}C) = 39.60, \ \delta(^{1}H) = 2.55]$. Positive values of chemical shifts denote high frequency shifts with respect to standards.

Two-dimensional H,H-COSY, NOESY, gs (gradient selected)-HMQC and gs-HMBC [11,12] techniques were measured using standard micro-programs provided by Bruker.

2.3. Mass spectrometry

Mass spectra were measured on the ion trap analyser Esquire3000 (Bruker Daltonics, Bremen, Germany). The samples were dissolved in 50% aqueous acetonitrile and analysed by direct infusion at the flow rate 1–3 µl/min. Mass spectra were recorded in the range m/z 15–1000 using electrospray ionization (ESI) in the negative-ion mode. The isolation width of the precursor ion for MS/MS and MSⁿ measurements was m/z = 4 and the collision energy was in the range 0.6–1 V depending on the stability of individual ions.

3. Results and discussion

3.1. NMR spectra

The ²⁷Al, ¹⁵N, ¹³C and ¹H chemical shifts of complexes **1b–4b** as well as starting azo dyes **1a–4a** measured in DMSO-D₆ are collected in Tables 1–4.

The ¹H and ¹³C chemical shifts were assigned using H,H-COSY, gs (gradient selected)-HMQC and gs-HMBC [11,12] [optimised for ¹J(¹³C, H) ca 150 Hz and ³J(¹³C, H) ca 8 Hz]. A correlation of proton H(3') with carbon of COO group in HMBC spectra was a key information for the assignment of proton and carbon resonances C(1')–C(7').

Only one set of appropriate NMR signals was obtained for each of compounds **1b–4b** which means that both ligands in a molecule of complexes are equivalent, at least on the NMR time scale. On the contrary, Schetty and Kuster [13] observed three components having considerably different shade in cobalt analogue of compound **2b**.

Table 1 ¹H, ¹³C, ¹⁵N and ²⁷Al NMR chemical shifts of compounds 1a,b in DMSO- d_6

| | 1a | | 1b | |
|----------------|-----------------|----------------------|-----------------|---------------------|
| | $\delta(^{1}H)$ | $\delta(^{13}C)$ | $\delta(^{1}H)$ | δ(¹³ C) |
| 1 | _ | 137.62 | _ | 140.46 |
| 2 | _ | 150.15 | _ | 157.28 |
| 3 | 6.99 | 118.34 | 6.18 | 122.12 |
| 4 | 7.51 | 131.03 | 7.30 | 135.46 |
| 5 | - | 142.03 | - | 138.73 |
| 6 | 7.89 | 127.29 | 7.69 | 109.97 |
| 7 | - | 33.88 | - | 33.58 |
| 8 | 1.36 | 31.17 | 1.34 | 30.99 |
| 1′ | - | 148.45 | - | 149.32 |
| 2′ | _ | 133.52 | _ | 128.65 |
| 3′ | 7.98 | 130.77 | 8.12 | 131.22 |
| 4′ | 7.59 | 130.63 | 7.56 | 129.01 |
| 5′ | 7.63 | 130.89 | 7.69 | 131.90 |
| 6' | 7.84 | 115.21 | 7.81 | 120.32 |
| 7′ | _ | 167.83 | _ | 164.76 |
| Na | - | 80. 9 ^a | - | -34.0^{a} |
| N _b | _ | 129.5 ^{a,b} | _ | 99.5 ^{a,b} |
| Al | - | - | - | 0.5 ^c |

^a $\delta(^{15}N)$.

^b ¹⁵N labelled compound.

^c $\delta(^{27}\text{Al}).$

The coordination number of aluminium atom in compounds **1b–4b** can be determined by the measurement of NMR spectra of two nuclei directly involved in this process: nitrogen and aluminium.

²⁷Al NMR spectroscopy is a powerful tool for coordination studies [14-17]. The ²⁷Al chemical shifts are determined by two main factors: coordination number and the nature of ligands. The ²⁷Al chemical shifts move generally downfield passing from six- to five- and four-coordinated aluminium atoms, respectively. The ²⁷Al chemical shifts in compounds 1b-4b were found to resonate in a very narrow range from 0.5 to 2.3 ppm and they give the evidence for the aluminium central atom to be six-coordinated, because the ²⁷Al chemical shifts practically coincide with the signal of standard. Hence, two measurements had to be performed for each sample, one with the standard in a coaxial capillary, in which the absolute resonance frequency of standard was determined, and the second one, in which relatively broad ²⁷Al NMR signal of a complex was detected.

| | 5 | | | |
|----------------|-----------------|----------------------|--------------------------|---------------------|
| | 2a | | 2b | |
| | $\delta(^{1}H)$ | δ(¹³ C) | $\delta(^{1}\mathrm{H})$ | δ(¹³ C) |
| 1 | _ | 130.59 | _ | 128.78 |
| 2 | _ | 179.16 | _ | 160.40 |
| 3 | 6.67 | 126.87 | 6.42 | 124.51 |
| 4 | 7.88 | 142.71 | 7.80 | 139.90 |
| 4a | _ | 128.37 | _ | 127.06 |
| 5 | 7.68 | 129.32 | 7.80 | 128.13 |
| 6 | 7.46 | 127.18 | 7.47 | 124.43 |
| 7 | 7.59 | 129.36 | 7.69 | 128.32 |
| 8 | 8.41 | 122.22 | 8.56 | 121.28 |
| 8a | _ | 133.37 | _ | 133.90 |
| 1′ | — | 144.09 | - | 150.25 |
| 2' | — | 117.08 | _ | 134.33 |
| 3′ | 8.06 | 131.56 | 8.13 | 131.97 |
| 4′ | 7.33 | 124.50 | 7.54 | 128.78 |
| 5′ | 7.77 | 134.51 | 7.74 | 132.22 |
| 6' | 8.34 | 116.32 | 7.87 | 120.90 |
| 7′ | — | 167.60 | - | 165.21 |
| Na | 16.42 | -195.7 ^a | _ | $-59.7^{\rm a}$ |
| N _b | — | -21.4 ^{a,b} | - | 75.0 ^{a,b} |
| Al | _ | - | _ | 1.6 ^c |

Table 2 ¹H, ¹³C, ¹⁵N and ²⁷Al NMR chemical shifts of compounds **2a,b** in DMSO-*d*₆

^a $\delta(^{15}N)$.

^b ¹⁵N labelled compound.

^c δ(²⁷Al).

To decide if the sixfold coordination is due to solvent interaction (because DMSO is known to be a strong coordinating solvent) or to interaction with one nitrogen of each azo/hydrazone bond, additional indications come from ¹⁵N NMR spectra. In previous papers [7,9] we reported on the application of ¹⁵N NMR as a very sensitive technique for the detection of the existence of coordination as well for the determination of coordination position of nitrogens in -N=N- and -NH-N= groups, respectively. To assign ¹⁵N resonances undoubtedly and because of low solubility of compounds studied, it was necessary to prepare ¹⁵N selectively enriched compounds for the measurement of ¹⁵N chemical shifts.

Upfield ¹⁵N chemical shifts of about 100 ppm were observed on protonation of various heterocyclic derivatives, e.g. pyridine, purine, indolizine, etc. [18,19], where the nitrogen atom lone pairs are engaged in a bonding. The similar effect $[\Delta\delta(^{15}N) = 102-146 \text{ ppm}]$ [20] is also typical of the coordination of a nitrogen atom of azo dyes

Table 3 ¹H, ¹³C, ¹⁵N and ²⁷Al NMR chemical shifts of compounds **3a,b** in DMSO-*d*₆

| | 3a | | 3b | |
|----------------|-----------------|-----------------------|-----------------|-----------------------|
| | $\delta(^{1}H)$ | δ(¹³ C) | $\delta(^{1}H)$ | $\delta(^{13}C)$ |
| 1 | 2.54 | 26.20 | 2.61 | 26.30 |
| 2 | _ | 198.97 | _ | 198.61 |
| 3 | _ | 129.27 | _ | 125.48 |
| 4 | - | 160.69 | - | 160.06 |
| 5 | _ | 137.53 | _ | 135.87 |
| 6 | 7.67 | 120.28 | 7.17 | 120.71 |
| 7 | 7.39 | 129.15 | 7.07 | 128.83 |
| 8 | 7.17 | 124.53 | 7.07 | 125.10 |
| 1′ | - | 143.56 | - | 150.31 |
| 2′ | _ | 116.41 | _ | 125.88 |
| 3′ | 7.99 | 131.50 | 8.08 | 131.75 |
| 4′ | 7.22 | 123.69 | 7.34 | 125.20 |
| 5′ | 7.66 | 134.46 | 7.66 | 132.07 |
| 6′ | 7.92 | 115.28 | 7.73 | 120.13 |
| 7′ | - | 168.07 | - | 165.77 |
| Na | 14.99 | -211.3 ^{a,b} | _ | -107.9 ^a |
| N _b | _ | -12.5 ^{a,c} | _ | 62.2 ^{a,c} |
| N _c | 11.10 | -246.5 ^{a,d} | 12.48 | -251.0 ^{a,e} |
| Al | _ | - | _ | 1.0^{f} |

^a $\delta(^{15}N)$.

 $^{b-1}J(^{15}N, H) = 99.7 Hz.$

^c ¹⁵N labelled compound.

 $^{d-1}J(^{15}N, H) = 88.7 Hz.$

 e ¹ $J(^{15}N, H) = 88.7 Hz.$

^f $\delta(^{27}\text{Al}).$

provided the dyes exist in azo form. This is the case of compound **1b**. On the other hand, a coordination can cause a relatively large downfield shift in case a ligand exists in the hydrazone form (compounds **2b–4b**) and changes in δ ⁽¹⁵N) are due to a combination of two effects: the coordination (responsible for an upfield shift) and the change of the hydrazone tautomeric form to the azo form (responsible for even greater downfield shift). For details see Refs. [7,9].

We measured the ¹⁵N chemical shifts of compound **1b–4b**, using in ¹⁵N_b enriched (95% ¹⁵N) compounds using standard 1D spectra and both resonances at the natural abundance level using gs-¹H–¹⁵N correlation spectra. Having compared the data with cobalt(III) compounds [7] it is clear that very similar ¹⁵N chemical shifts were measured in compound **1b–4b** giving the evidence

| | 4a | | 4b | |
|----------------|-----------------|-----------------------|-----------------|---------------------|
| | $\delta(^{1}H)$ | δ(¹³ C) | $\delta(^{1}H)$ | δ(¹³ C) |
| 1 | _ | -194.2ª | _ | |
| 2 | _ | -74.3ª | _ | -96.2ª |
| 3 | _ | 148.60 | _ | 150.24 |
| 4 | _ | 129.79 | - | 127.82 |
| 5 | _ | 155.81 | _ | 151.33 |
| 6 | _ | 137.99 | - | 137.16 |
| 7 | 7.90 | 117.70 | 7.31 | 119.18 |
| 8 | 7.44 | 128.95 | 7.20 | 128.88 |
| 9 | 7.21 | 124.72 | 7.17 | 125.90 |
| 10 | 2.29 | 11.71 | 2.46 | 11.80 |
| 1′ | _ | 143.15 | - | 149.38 |
| 2′ | - | 116.19 | _ | 127.32 |
| 3′ | 7.99 | 131.42 | 8.04 | 131.88 |
| 4′ | 7.23 | 124.27 | 7.46 | 127.27 |
| 5′ | 7.66 | 134.40 | 7.54 | 128.78 |
| 6' | 7.92 | 115.02 | 7.85 | 120.21 |
| 7′ | _ | 167.95 | _ | 165.45 |
| Na | 14.61 | -209.3 ^{a,b} | _ | -68.9^{a} |
| N _b | _ | -24.7 ^{a,c} | _ | 58.1 ^{a,c} |
| A 1 | _ | _ | _ | 2 3d |

Table 4 ¹H. ¹³C. ¹⁵]

| H, ¹³ C, ¹⁵ N and ²⁷ Al NMR | chemical shifts | of compounds 4a,b |
|--|-----------------|-------------------|
| n DMSO-de | | |

^a $\delta(^{15}N)$.

^{b 1} $J(^{15}N, H) = 100.3$ Hz.

^{c 15}N labelled compound.

^d δ(²⁷Al).

of N_a nitrogen coordination [7,9] with aluminium atom.

The conclusion is supported by Jagi's observation [21] who performed an X-ray study of cobalt complex of a dye, its ligand being derived from anthranilic acid, and also found the cordination of cobalt atom with nitrogen N_a .

The mutual differences in ¹H, ¹³C and ¹⁵N chemical shifts in compounds **1b–4b** are slightly greater then those in analogous hydroxy derivatives [9]. The reason might be that coordination procedure provides two new six-membered rings (see Scheme 1) while in analogous compounds in which –COO-group is substituted by –O-group [9] new six-membered and five-membered rings exist as a consequence of coordination. Due to a greater flexibility of six membered rings, small changes in bond lengths and bond angles can be expected.





Scheme 3.

3.2. Mass spectra

The complexes of three-valent metals with azo compounds carry a negative charge themselves, which is ideally suited for the negative-ion electrospray ionization (ESI) [22]. Similarly to the complexes of Al(III), Co(III) and Fe(III) with o,o'dihydroxyazodyes, the molecular anionic species [M-cation]⁻ are the only ions in the mass spectra of Al(III) complexes with $o_{,o'}$ -hydroxycarboxyazodyes (see Scheme 3a), which enables an easy molecular weight determination of all studied compounds. The character of cation does not influence ESI mass spectra in the negative-ion mode. The background noise in the spectra is very low, therefore ESI mass spectra can be used for monitoring of purity of synthesised complexes. The MS/MS analysis with the ion trap analyser is suitable for confirmation of expected structures, as demonstrated recently [23]. Scheme 3b shows an MS/MS spectrum of precursor ion at m/z 667, which confirms the structure assignment of compound 4b, as shown on inset fragmentation scheme. The main observed losses are CO_2 (m/z 44), C₆H₃ (m/z 75), CO₂ and CH₃CN (m/z 85), phenylisocyanate $(m/z \ 119)$ and N₂ $(m/z \ 28)$. The typical feature of MS/MS spectra is the subsequent loss of the same fragments for both organic ligands, e.g. 85 (667-582 and 463-378), 75 (582-507 and 378-303) and 44 (507-463 and 303-259). This approach was applied for the structure verification of all studied complexes.

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